

Does telomere elongation in cloned organisms lead to a longer lifespan if cancer is considered?

Michael Masa¹, Stanisław Cebrat² and Dietrich Stauffer¹

¹ Institute for Theoretical Physics, Cologne University, D-50923 Köln, Euroland ² Institute of Genetics and Microbiology, University of Wrocław, ul. Przybyszewskiego 63/77, PL-54148 Wrocław, Poland

Abstract

As cell proliferation is limited due to the loss of telomere repeats in DNA of normal somatic cells during division, telomere attrition can possibly play an important role in determining the maximum life span of an organism. With computer simulations of cell culture development in organisms, which consist of tissues of normal somatic cells with finite growth, we obtain an increase of life span for longer telomeric DNA in the zygote. By additionally considering a two-mutation model for carcinogenesis and indefinite proliferation by the activation of telomerase, we demonstrate that the risk of dying due to cancer can outweigh the positive effect of longer telomeres on the longevity.

Keywords: Biological Ageing; Computer simulations; Telomeres; Telomerase; Cancer

1 Introduction

Telomeres are tandem repeated noncoding sequences of nucleotides at both ends of the DNA in eukaryotic chromosomes stabilizing the chromosome ends and preventing them from end-to-end fusion or degradation [1]. Polymerase cannot completely replicate the 3' end of linear DNA, so telomeres are shortened at each DNA replication [2]. This end replication problem leads to a finite replicative capacity for normal somatic cells [3]. They can only divide up to a certain threshold, the Hayflick limit [4]. The enzyme telomerase, repressed in most normal somatic cells, synthesizes and elongates telomere repeat sequences at the end of DNA strands so that certain cells like germline cells are immortal and indefinite in growth [5, 6].

Most forms of cancer follow from the accumulation of somatic mutations [7]. Cancer-derived cell lines and 85-90% of primary human cancers are able to synthesize high levels of telomerase and thus are able to prevent further shortening of their telomeres and proliferate indefinitely [8]. But if cells are premalignant or already cancerous and telomerase is not yet activated, the proliferation of these cells and the accumulation of mutations is determined by their further replicative capacity [9]. So the frequency of malignant cancer should be higher for longer telomeres.

Recently published data show that longer telomeric DNA increased the life span of nematode worms [10]. So there may be a positive effect on the longevity of complex cloned organisms with renewing tissues if the telomere length in zygote cells is increased [11]. As the probability for the incidence of cancer is correlated with the replicative potential of the mutated cells [12], one can ask the following

question: Is an extension of lifespan of cloned organisms possible if telomeres in embryonic cells are elongated and cancer is considered? An answer to this question could be given by computer simulations as the presented model focuses on ageing by the loss of telomeres in DNA.

After presenting the basic model of telomere attrition in organisms and computation results, we explain how cancer and telomerase are introduced and discuss the effects of different initial telomere lengths.

2 Basic model of biological ageing due to telomere shortening

As shortening of telomeres is one of the supposed mechanisms of ageing on cellular level, there are many different approaches to model telomere loss [13, 14, 15, 16]. In our basic model every organism is developed from a single progenitor cell, the zygote (figure 1). The initial telomere length of zygote cells is assumed to be normally distributed with mean μ_z and standard deviation σ_z [17]. Telomere repeats lost per division (TRLPD) are randomly chosen at each division of every cell from a normal distribution with mean μ_{TRLPD} and standard deviation σ_{TRLPD} . A dividing cell produces a clone who inherits the replicative capacity of the progenitor cell at this age. All normally distributed variables are generated with the Box-Muller method [18]. Cells can divide until nearly all their original telomeres are lost.

For every organism the dynamics of the model is as follows: Divisions of the zygote and the stem cells derived from it occur 6 times in the early embryo. Each of these cells is the progenitor of one tissue. This is followed by a period of population doublings where all cells divide once in every timestep until 2^7 cells are present in each tissue. In the following maturation stage, cells are chosen randomly for division until each tissue reaches the adult size of 10^4 cells. It takes about 26 timesteps until an organism is mature.

Ageing starts now, cells first die with 10% probability due to events like necrosis or apoptosis. 10% of the cells of the corresponding tissue are then randomly chosen for division to fill this gap. The replacement does not have to be complete as the chosen cell could probably not divide anymore due to telomere attrition. After some time the tissue will start shrinking. The random choice of dying and dividing cells in differentiated tissues is in accordance with nature as for example in epithelium the choice of cells to be exported from the basal layer is random [19, 20]. The organism dies, if its actual size reaches 50% of its mature size.

2.1 Results without cancer

Age distributions for different mean telomere lengths in the zygote cells are shown (figure 2). The shape of these distributions is very analogous to empirical data of many human and animal populations. We also obtain a positive effect on the longevity of the organisms if the mean telomere length in the precursor cell is increased.

The chosen mean doubling potentials for primary cells are 30, 40 and 50 with the choice of $\mu_z = 1500, 2000, 2500$ and $\mu_{TRLPD} = 50$. The number of mitotic divisions

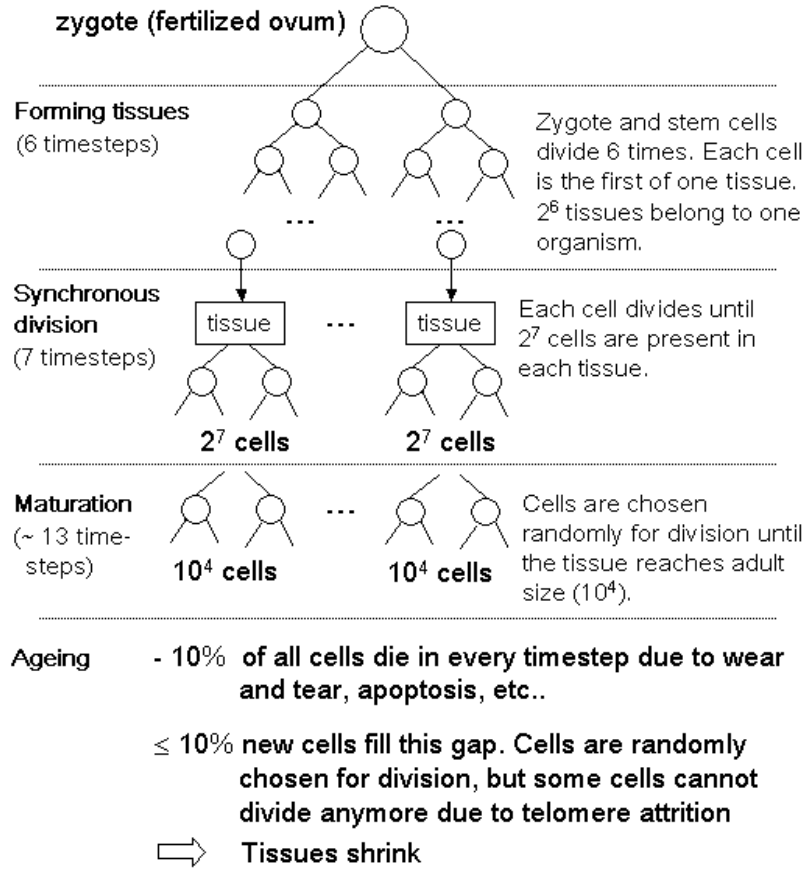


Figure 1: Dynamics of cell proliferation in one organism in the basic model

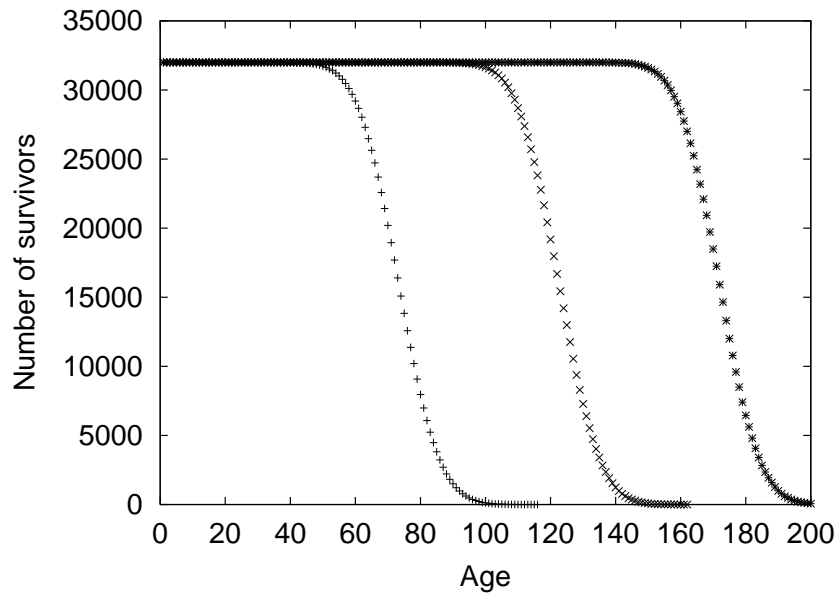


Figure 2: Age distribution of 32000 organisms with telomere lengths of $\mu_z = 1500(+)$, $\mu_z = 2000(\times)$ and $\mu_z = 2500(*)$; $\sigma_z = 100$, $\mu_{TRLPD} = 50$, $\sigma_{TRLPD} = 10$

observed in human fibroblasts is higher [21], but the choice of this parameters is reasonable as the number of considered cells per mature organism (640000) in this model is also much lower than in human organisms where the total number of cells is of the order of 10^{13} [19].

3 Introducing carcinogenesis and telomerase

Clonal cancer is now introduced in the model. In accordance to the model of Moolgavkar et al., one of our assumptions is that malignant tumors arise from independently mutated progenitor cells [22]. For most forms of carcinoma, transformation of a susceptible stem cell into a cancer cell is suggested to be a multistage process of successive mutations with a relatively low probability for the sequential stages [7, 23]. Two independent and irreversible hereditary mutation stages are considered here, which can occur at every level of development of the organism during cell division.

The first premalignant stage is a promotion: A dividing cell can mutate with small probability p_{mut} [19]. All descendant cells inherit this mutation. This mutation leads to a partial escape from homeostatic control of growth by the local cellular environment [9, 24]. Cells on the promotion stage have a selective advantage over unaffected cells [25]. In our model they are chosen first for division during maturation and for filling up the gap in the ageing period.

The subsequent transition can occur again with probability p_{mut} during division. If a cell reaches this second stage of mutation it is a progenitor of a carcinoma. An explosive clonal expansion to a fully malignant compartment happens [24]. This cell and the clonal progeny doubles in the current timestep until it is no more possible due to telomere attrition. This expansion leads only to an increase of the malignant cell population size. As a certain fraction of cells is killed per unit time and clonal expansions only occur with a very small probability, the tumor environments may not continue growing, eventually shrink, or even die [26]. We assume that it is necessary for advanced cancer progression and therefore for the development of a deadly tumor that fully mutated cells are able to activate telomerase [27].

In our model, telomerase activation is possible at every age of the organism in normal and mutated cells during division with a very low probability p_{telo} . The irreversible loss of replicative potential is stopped in these cells. As the contribution of telomerase to tumorigenicity is not yet completely understood, [28, 29, 30], we assume that death of an organism due to cancer occurs if telomerase is reactivated in at least one fully mutated cell [31]. We treat the time interval between the occurrence of a deadly tumor and death due to it as constant, so we set this interval to zero. In our carcinogenic process there are two ways to reach the critical stage of indefinite proliferation of immortal cancerous cells:

1. Telomerase is already reactivated in a cell which reaches the second mutation stage.
2. Activation of telomerase occurs in one fully mutated cell during the explosive clonal expansion of cancerous cells after a cell has reached the second stage of mutation.

3.1 Effects of different telomere lengths considering cancer

Figure 3 shows simulation results for $\mu_z = 1500$ and $\mu_z = 2500$. As we considered

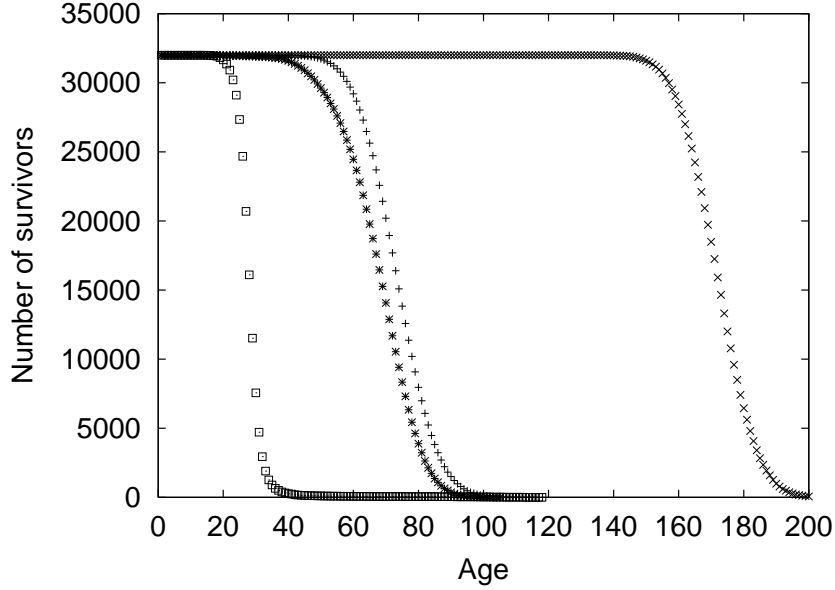


Figure 3: Age distribution of 32000 organisms with telomere lengths of $\mu_z = 1500$ with (*) and without cancer (+) and $\mu_z = 2500$ with (□) and without cancer (×); $\sigma_z = 100$, $\mu_{TRLPD} = 50$, $\sigma_{TRLPD} = 10$. Cancer mutations are possible with $p_{mut} = 5 * 10^{-5}$. Telomerase can be activated with $p_{telo} = 10^{-5}$.

a lower complexity by choosing a lower number of tissues and cells per organism, we assumed higher mutation rates for the incidence of cancer than observed in nature [32, 33].

The age distribution for shorter initial telomere lengths considering cancer is shifted to the left but still very old organisms exist. Cancer increases only the probability to die in a certain range of age which varies for different telomere lengths. For longer telomeres the age distribution is again shifted to the left but even behind the distributions for shorter telomeres with and without considering cancer. There is a strong increase in the risk of dying because of cancer and hence the life expectation is much lower than for shorter initial telomeres.

The force of mortality resulting from this model is shown for $\mu_z = 1500$ with and without considering cancer in comparison to empirical human mortality data (figure 4). It agrees very well with human mortality functions provided cancer is incorporated into the model, and shows a mortality deceleration at advanced age,

4 Conclusion

The expected simulation result of the basic model without cancer is an increase of life span of most organisms with longer initial telomeres. After introducing somatic mutations promoting cancer and telomerase activation in this model, the survival probability is lower for each considered initial telomere length in certain time intervals in adult ages.

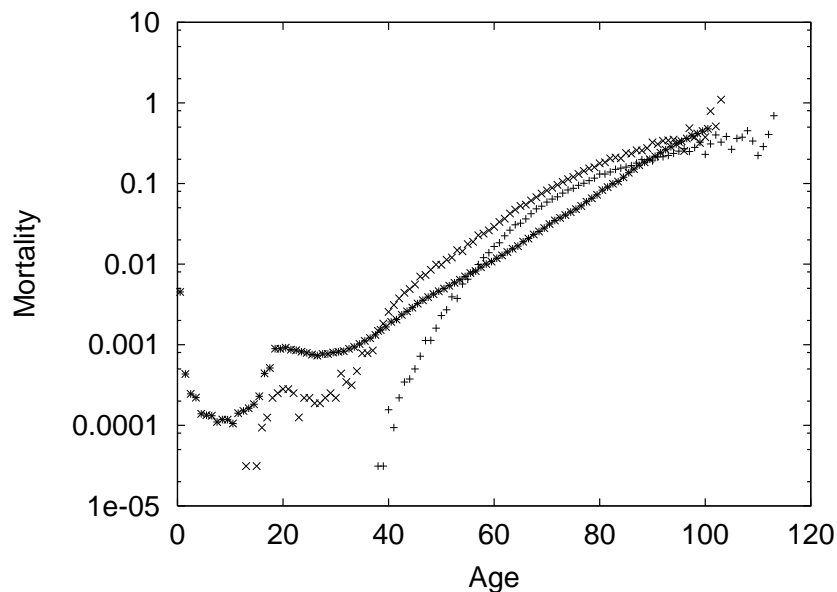


Figure 4: Mortality function for $\mu_z = 1500$ with (\times) and without cancer ($+$), 32000 organisms considered, $\sigma_z = 100$, $\mu_{TRLPD} = 50$, $\sigma_{TRLPD} = 10$; German men ($*$) from www.destatis.de (June 2004) (Sterbetafel 2000/2002)

But even low probabilities for the two mutation stages and for the activation of telomerase lead to a strong reduction of life span for longer telomeres. So the implication of two-stage carcinogenesis for the incidence of cancer in this simple model of cell proliferation in organisms is that the life span of complex cloned organisms cannot be increased by artificially elongating telomeres in primary cells.

Further improvements, extensions and applications of this model are possible. With respect to the role of telomeres and telomerase in carcinogenesis, maybe this computational approach can contribute to the development of a comprehensive theoretical model in oncology uniting mutagenesis and cell proliferation [34].

Acknowledgements

We wish to thank the European project COST-P10 for supporting visits of MM and DS to the Cebrat group at Wrocław University and the Jülich supercomputer center for computing time on their CrayT3E. CS was supported by Foundation for Polish Science.

References

- [1] Blackburn EH. Structure and function of telomeres. *Nature* 1991 Apr;350:569-72.
- [2] Harley CB, Futcher AB, Greider CW. Telomeres shorten during ageing of human fibroblasts. *Nature* 1990 May;345:458-60.
- [3] Olovnikov AM. Theory of marginotomy. *J Theor Biol* 1973 Sep;41:181-90. er and dying in the attempt

- [4] Hayflick L. Living forever and dying in the attempt. *Exp Gerontology* 2003 Jun;38:1231-41.
- [5] Greider CW, Blackburn EH. Identification of a specific telomere terminal transferase activity in *Tetrahymena* extracts. *Cell* 1985 Dec;43(2 Pt 1):405-13.
- [6] Shay JW, Wright WE. Hayflick, his limit, and cellular ageing. *Nature Rev* 2000 Oct;1:72-6.
- [7] Nordling CO. A new theory on the cancer inducing mechanism. *Br J Cancer* 1953 Mar;7(1):68-72.
- [8] Kim NW, Piatyszek MA, Prowse KR, Harley CB, West MD, Ho PL, Coviello GM, Wright WE, Weinrich SL, Shay JW. Specific association of human telomerase activity with immortal cells and cancer. *Science* 1994 Dec 23;266(5193):2011-5.
- [9] Moolgavkar SH, Luebeck EG. Multistage carcinogenesis and the incidence of human cancer. *Genes Chromosomes Cancer* 2003 Dec;38(4):302-6.
- [10] Joeng KS, Song EJ, Lee KJ, Lee J. Long lifespan in worms with long telomeric DNA. *Nat Genet* 2004 Jun;36(6):607-11. Epub 2004 May 02.
- [11] Lanza RP, Cibelli JB, Blackwell C, Cristofalo VJ, Francis MK, Baerlocher GM, Mak J, Schertzer M, Chavez EA, Sawyer N, Lansdorp PM, West MD. Extension of cell life-span and telomere length in animals cloned from senescent somatic cells. *Science* 2000 Apr 28;288(5466):665-9.
- [12] Shay JW, Wright WE. Telomeres and telomerase: implications for cancer and aging. *Radiat Res* 2001 Jan;155(1 Pt 2):188-193.
- [13] Levy MZ, Allsopp RC, Futcher AB, Greider CW, Harley CB. Telomere end-replication problem and cell aging. *J Mol Biol* 1992 Jun 20;225(4):951-60.
- [14] Aviv A, Levy D, Mangel M. Growth, telomere dynamics and successful and unsuccessful human aging. *Mech Ageing Dev* 2003 Jul;124(7):829-37.
- [15] Olofsson P, Kimmel M. Stochastic models of telomere shortening. *Math Biosci* 1999 Apr;158(1):75-92.
- [16] Sozou PD, Kirkwood TB. A stochastic model of cell replicative senescence based on telomere shortening, oxidative stress, and somatic mutations in nuclear and mitochondrial DNA. *J Theor Biol* 2001 Dec 21;213(4):573-86.
- [17] op den Buijs J, van den Bosch PP, Musters MW, van Riel NA. Mathematical modeling confirms the length-dependency of telomere shortening. *Mech Ageing Dev* 2004 Jun;125(6):437-44.
- [18] Chinellato O. www.wr.inf.ethz.ch/education/pr/files/u8/bm.pdf
- [19] Cairns J. Mutation selection and the natural history of cancer. *Nature* 1975 May 15;255(5505):197-200.

- [20] Frank SA, Iwasa Y, Nowak MA. Patterns of cell division and the risk of cancer. *Genetics* 2003 Apr;163(4):1527-32.
- [21] Allsopp RC, Vaziri H, Patterson C, Goldstein S, Younglai EV, Futcher AB, Greider CW, Harley CB. Telomere length predicts replicative capacity of human fibroblasts. *Proc Natl Acad Sci USA* 1992 Nov 1;89(21):10114-8.
- [22] Moolgavkar SH, Dewanji A, Venzon DJ. A stochastic two-stage model for cancer risk assessment. I. The hazard function and the probability of tumor. *Risk Anal* 1988 Sep;8(3):383-92. Related Articles, Links
- [23] Armitage P, Doll R. The age distribution of cancer and a multi-stage theory of carcinogenesis. *Br J Cancer* 1954 Mar;8(1):1-12.
- [24] Sarasin A. An overview of the mechanisms of mutagenesis and carcinogenesis. *Mutat Res* 2003 Nov;544(2-3):99-106.
- [25] Armitage P, Doll R. A two-stage theory of carcinogenesis in relation to the age distribution of human cancer. *Br J Cancer* 1957 Jun;11(2):161-9.
- [26] Hiyama E, Hiyama K, Yokoyama T, Matsuura Y, Piatyszek MA, Shay JW. Correlating telomerase activity levels with human neuroblastoma outcomes. *Nat Med* 1995 Mar;1(3):249-55.
- [27] Shay JW. Toward identifying a cellular determinant of telomerase repression. *J Natl Cancer Inst* 1999 Jan 6;91(1):4-6.
- [28] Hiyama T, Yokozaki H, Kitadai Y, Haruma K, Yasui W, Kajiyama G, Tahara E. Overexpression of human telomerase RNA is an early event in oesophageal carcinogenesis. *Virchows Arch* 1999 Jun;434(6):483-7.
- [29] Kanjuh V, Knežević M, Živka E, Ostojić M, Beleslin B. Conceptions about cancer genetics and immortality of cancer cells in 2000. *Archive of Oncology* 2001 9(1):3-4
- [30] Blasco MA, Hahn WC. Evolving views of telomerase and cancer. *Trends Cell Biol* 2003 Jun;13(6):289-94.
- [31] Moolgavkar SH, Knudson AG Jr. Mutation and cancer: a model for human carcinogenesis. *J Natl Cancer Inst* 1981 Jun;66(6):1037-52.
- [32] Luebeck EG, Moolgavkar SH. Multistage carcinogenesis and the incidence of colorectal cancer. *Proc Natl Acad Sci USA* 2002 Nov 12;99(23):15095-100. Epub 2002 Nov 01.
- [33] Drake JW, Charlesworth B, Charlesworth D, Crow JF. Rates of spontaneous mutation. *Genetics* 1998 Apr;148(4):1667-86.
- [34] Gatenby RA, Maini PK. Mathematical oncology: cancer summed up. *Nature* 2003 Jan 23;421(6921):321.